

# Leica SR GSD 3D TIRF (Irchel)

How to start up and mount a sample on the Leica SR GSD 3D TIRF microscope.

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# INTRODUCTION

In this guide of the Center for Microscopy and Image Analysis we show how to start up and mount a sample on the Leica SR GSD 3D TIRF microscope.

Please find more information about the system here.

# Step 1 — Microscope hardware



- Laser Rack
- Microscope body ("DMI6000")
- Microscope control box ("CTR box")
- Fluorescence lamp
- Isolation Table
- Computer

## Step 2 — Switch ON microscope



(i) The control box is always kept ON to maintain steady temperature of the microscope components.

- Switch OFF the CTR box, as lasers must be turned ON BEFORE the CTR box.
- At the laser rack:
  - **Turn ON** the main power switch.
  - Turn the laser key for the 405 nm laser to "I" in order to enable "LASER Emission".
  - Switch ON all other lasers. Do NOT turn the keys yet.
  - The LEDs ("SHG Ready") light shortly up in green and turn then orange.
  - Once continuously green, turn laser keys to "LASER ON".

## Step 3 — CTR box, EL6000 and isolation table



- Switch ON the control box.
- Switch ON the fluorescence lamp.

⚠ Once turned on, the lamp should stay on for at least 30 min.

- Switch ON the isolation table.
  - Press "E" in order to enable isolation (indicated by the red LED "ISOL. ON" and display).

#### Step 4 — Turn on computer and sign-in



- Turn ON the computer.
- Sign-in with your ZMB core credentials.

#### Step 5 — Start the "LAS X" software



- Start the "LAS X" software via the desktop icon.
- Stop the countdown by clicking on it.
- Make sure "DefaultDynamicWidefieldTree.xlhw" and "Standard Configuration" is selected.
- Click "OK".

# Step 6 — Switch ON the lasers

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- Make sure you are in the "Main" module.
- Go to "Configuration".
- Select "Laser Config".
- Switch "ON" the lasers you need.
  - The "LASER ENABLED" light should turn white.
- Go back to "Acquire".

## Step 7 — 160x / 1.43NA oil objective



• Select the **160x/1.43** objective in the "LAS X" software.

(i) The **160x/1.43** objective is the only suitable objective for **GSD imaging**.

- Double-check if the objective is correctly inserted (clicked in).
- (i) The GSD objective is directly connected to the manual stage (SuMo-Stage) in order to reduce drift.

## Step 8 — Mount a sample



- **Push** the condenser arm to the **back**.
- Apply "Type F" immersion to your sample or onto the objective (do not touch the lens with the applicator).
- Insert a sample with the **coverslip facing down** and fix it with the two springs.
- Move your sample over the objective by using the manual stage knobs.
  - Movement in <u>y-direction</u>: *clockwise moves the stage towards the back.*
  - Movement in <u>x-direction</u>: *clockwise moves the stage to the right.*
- Pull back the condenser arm.

## Step 9 — Laser protection cover



- (i) An interlocked "Laser Protection Cover" is attached to the condenser arm to protect from the strong laser illumination needed for imaging.
- This cover has to be **opened** for accessing the sample.
- For engaging the lasers during imaging the **cover** has to be **closed**.
- **Proper closure** is confirmed by a green LED on the back of the microscope.

## Step 10 — Focus your sample in epi-fluorescence mode



- In the "LAS X" software:
  - Choose **FLUO** to enable widefield illumination.
  - Choose an **appropriate filter cube** for your fluorophore.
  - Set the camera **exposure time**.
- Click "Live".
- Use the external controller to focus your sample (max. travel range 400 um).

## Step 11 — Focus your sample - continued



- Switch to "TIRF" mode to enable laser illumination.
- Click "Live".
- Adapt laser power while being in "Live" mode.

## Step 12 — Starting the "GSD" wizard



- After finding the focus and proper laser and exposure settings switch to the "GSD" wizard.
- GSD operation will be explained in another ZMB guide.